

BMG 744
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Mass spectrometry imaging

Stephen Barnes, PhD

With sincere acknowledgments to David Stella, PhD and Kyle A. Floyd, MS, former students in the Barnes Laboratory (2005-2012) and Kevin Schey, PhD, Vanderbilt University

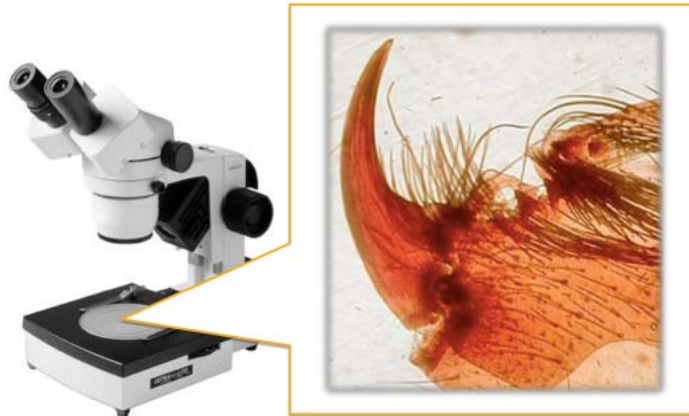
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What does MS imaging offer?

- **Can provide information that “grind and find” cannot**
 - **What is in the imaged section?**
 - **Where is it?**
 - **How much of it is there?**
 - **Is it modified?**
- **As we’ll see, much as the laser is targeted at the frozen section, it is an untargeted assay**

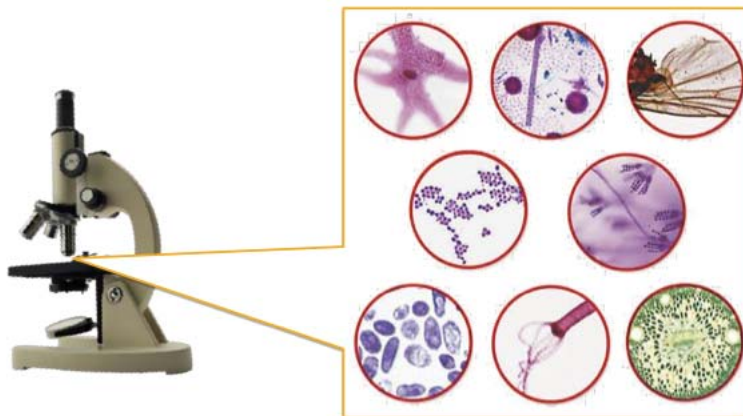
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Imaging is widely used in research



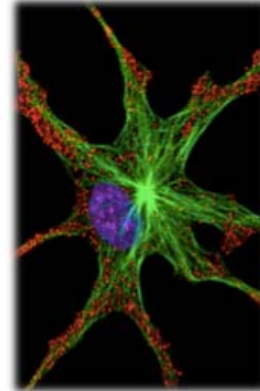
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Light microscopy



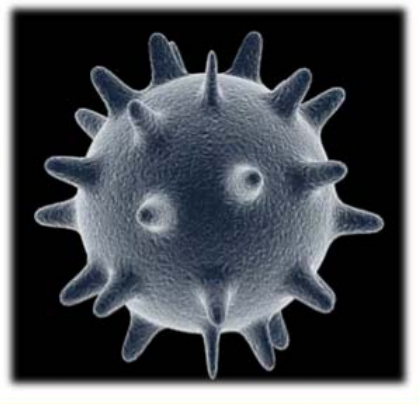
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Confocal Microscopy



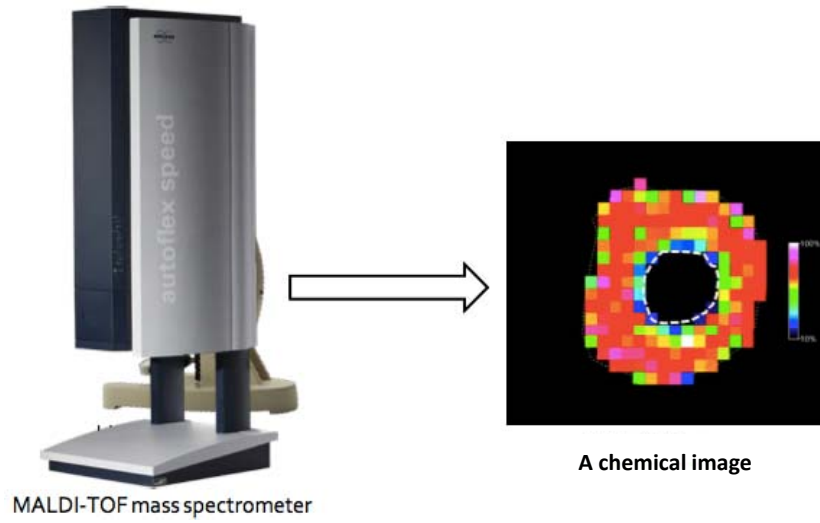
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Electron microscopy

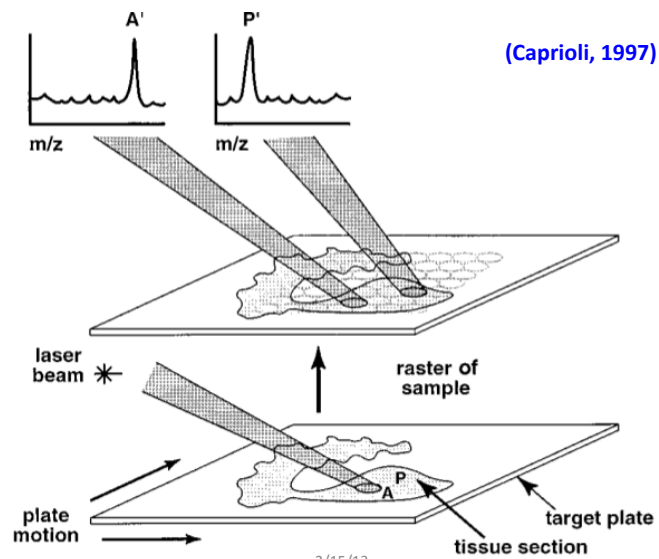


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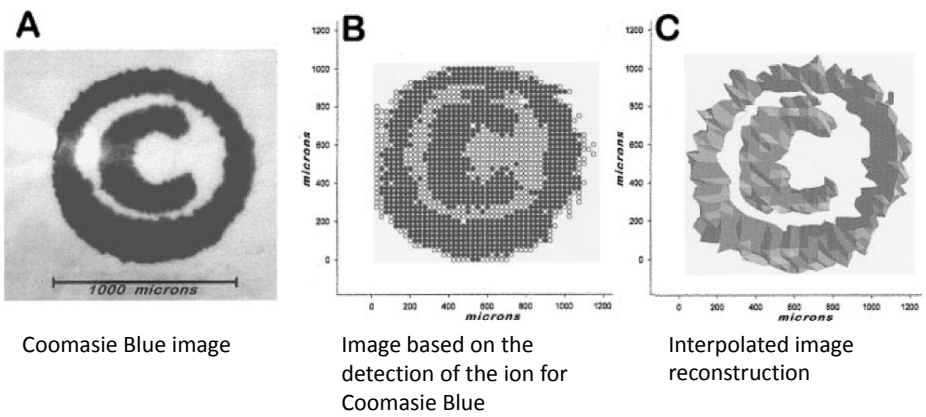
Mass spectrometry imaging



Principle of MALDI imaging



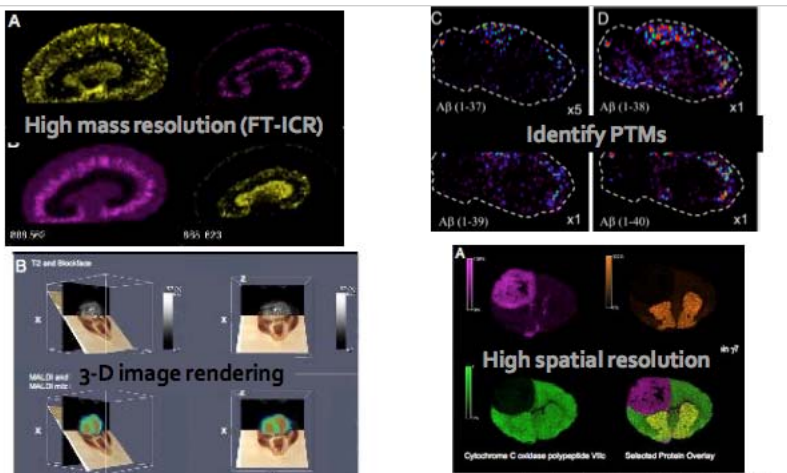
A simple example of MS imaging



(Caprioli, 1997)

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Modern applications of MS imaging



Seeley et al. (2008) PNAS:18126-31.

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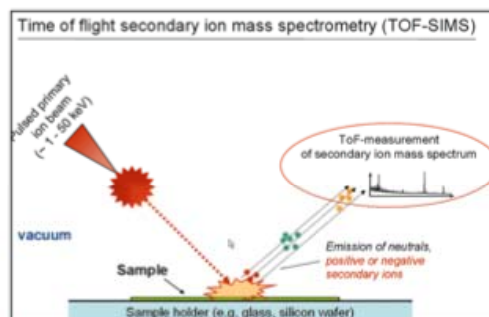
How to get analytes “off” a tissue section

- **Several types of ionization sources are used:**
 - **SIMS** – Secondary Ion MS
 - **DESI** – Desorption electrospray ionization
 - **MALDI**
 - **MALDESI**
 - **LAESI** – Laser Ablation Electrospray Ionization

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SIMS

- Used in early studies (elemental analyses)
- Very high resolution (>50 nm)
- Principle of ionization: collated ion beam

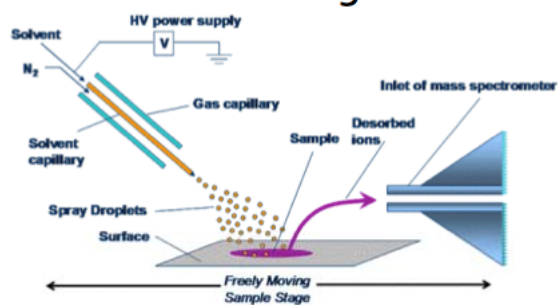


- Destructive, penetrating, low mass range

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DESI

- Moderate resolution (20 - 300 μm)
- Principle of ionization: Charged solvent spray

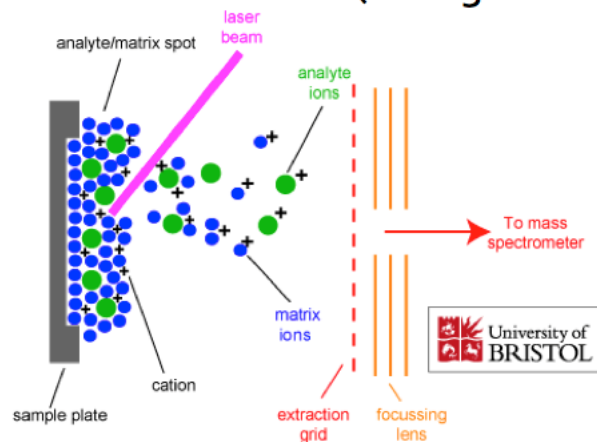


- Surface molecules, multiply charged, low - mid mass range

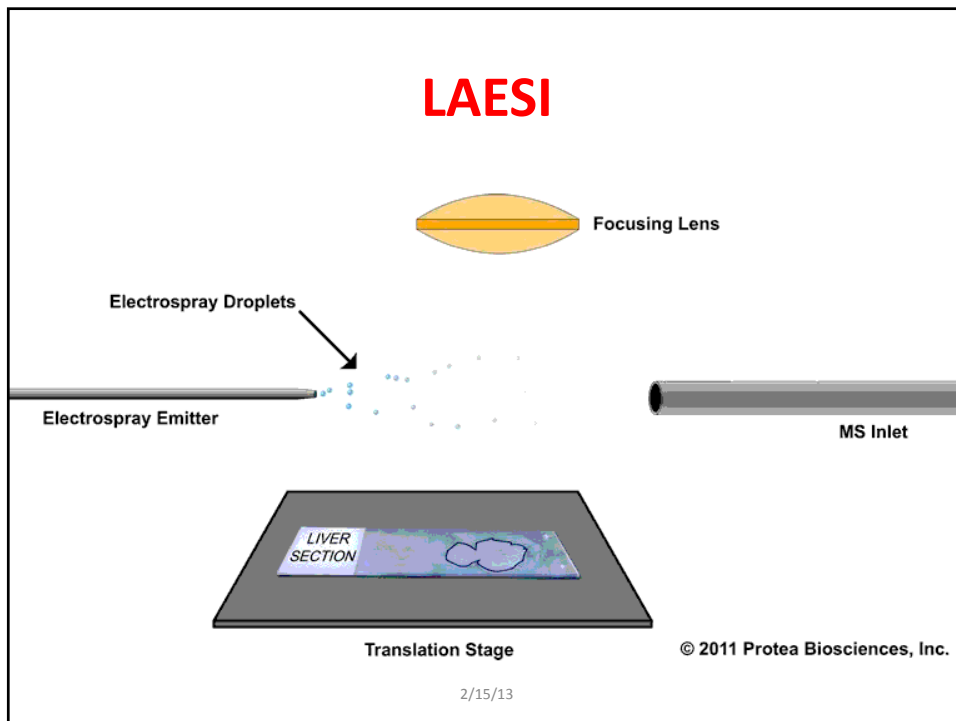
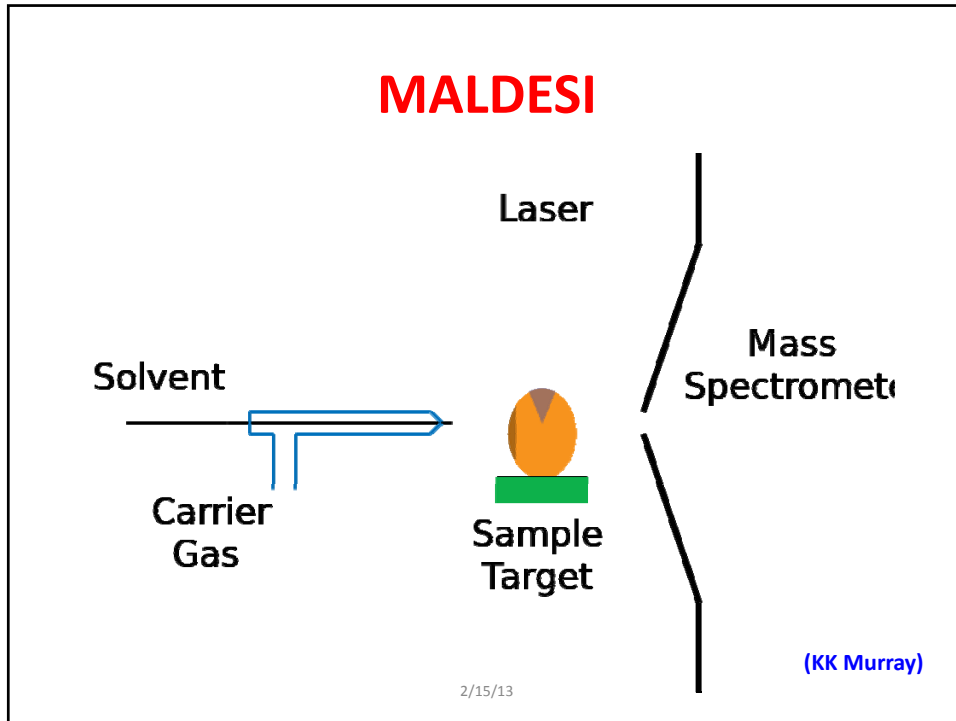
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MALDI

- Currently the more commonly used ionization source in MSI (biological research)



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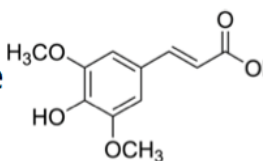
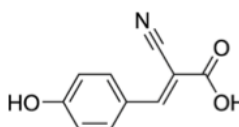
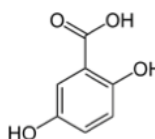
“Profiling” versus “imaging”

- **Profiling:**
 - Limited, directed information
 - Rapid analysis high throughput
 - Clinical applications and biomarker discovery
- **Imaging:**
 - Extensive, high resolution
 - Time consuming, laborious
 - Useful for investigative research

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MALDI matrices

- **DHB- 2,5-dihydroxybenzoic acid**
 - Commonly used for small molecules
- **CHCA- α -Cyano-4-hydroxycinnamic acid**
 - Commonly used for peptides and small proteins
- **SA- Sinapinic acid**
 - Commonly used for peptides and whole proteins (<100 kDa)



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Matrix solvent

- Acetonitrile is the solvent of choice
 - Range of percentage of solvent
 - Depends on application (tissue types)
 - Sometimes the solution is augmented with different additives including detergents
- Acid is also present in the matrix solution
 - Commonly formic acid
 - Promotes ionization
 - Ranges of percentage is also possible (upwards of 10%)

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How to spot the matrix

- Multiple technologies available:
 - Hand-spotting
 - TLC spraying
 - Sublimation
 - Precision mechanical spotting:
 - Acoustic devices.
 - Chemical printers
 - Inkjet printers

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Matrix application I: Manual spotting

✓ Start here!

- Gold standard for signal
- Best extraction
- Biggest crystals
- Best S/N and most signals
- Use for parameter optimization

× But don't use for imaging

- Poor resolution
- Limited spot placement accuracy
- Many cell types extracted together

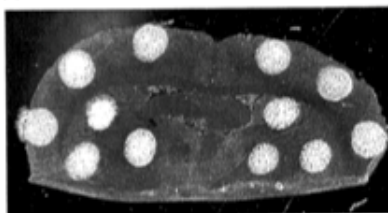


pipette
~250 nL
~1-2 mm diameter



pulled capillary
~100 nL
~800 μ m – 1 mm diameter

- Rat brain tissue
- SA
- 20 mg/ml
- 50:50 ACN:H₂O
- 0.25 μ l x 2



zoom in



AIMS 2012 Vanderbilt U

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Matrix application III: Manual coating

- TLC reagent sprayer
 - ✓ Commonly used
 - ✓ Variable reservoir sizes (10-25 ml)
 - ✓ Inert
 - × Inconsistent droplets from sprayer to sprayer



WS
#2A/5A

- Artist airbrush
 - ✓ Fine droplets
 - × May corrode



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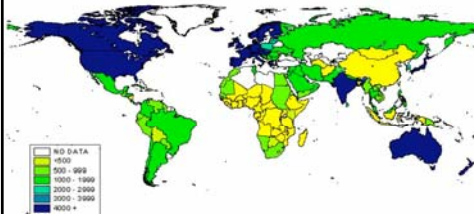
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Cataract disease in an Amazonian



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Cataract Disease and Public Health:

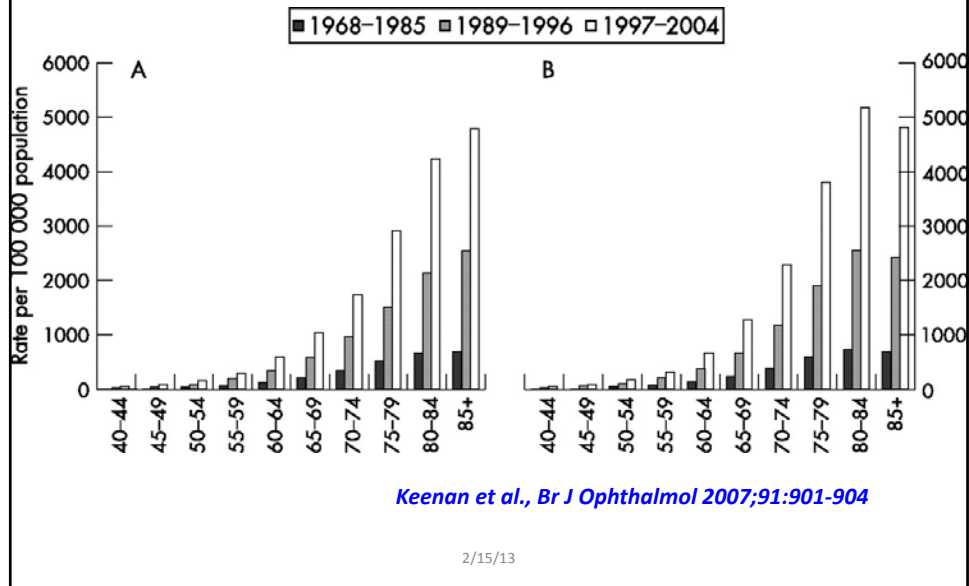


- WHO states that age-related cataract is responsible for 48% (~22 million people affected) of blindness in the world.
- Today people are living longer, putting more elderly at risk for cataract disease.
- Currently surgical removal is the only treatment, for which many countries have inadequate resources.
- In 2004 U.S. alone, cataract related medical expenses (i.e., surgical lens replacement) were estimated at \$6.8 billion dollars, or ~42% of total vision related costs. (Rein et al. 2006).
- Most common surgery in USA (431,000)

Map from WHO: http://www.who.int/blindness/data_maps/CSR_WORLD_2004.jpg
 Picture from WHO: <http://www.who.int/blindness/causes/priority/en/index1.html>

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Lens cataract incidence with aging



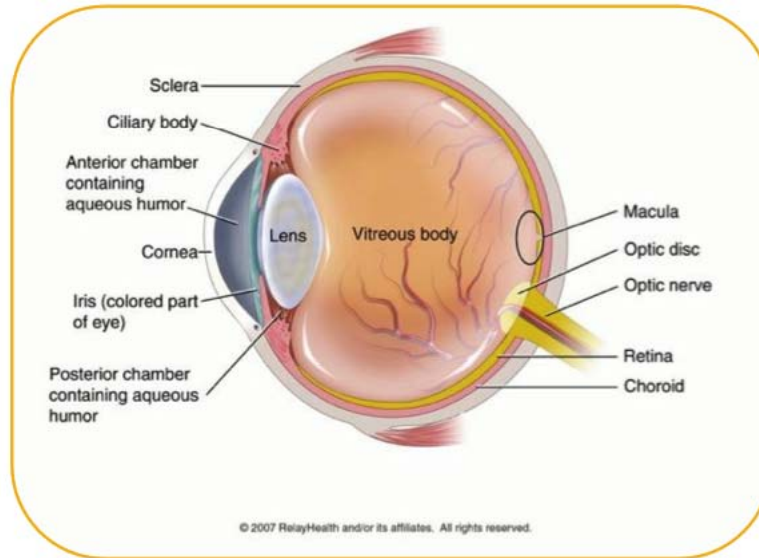
Lens changes occur in pets, too



Cloudiness is a natural event that occurs with aging and precedes cataract formation. Due to packing of the lens fiber cells.

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Anatomy of the eye



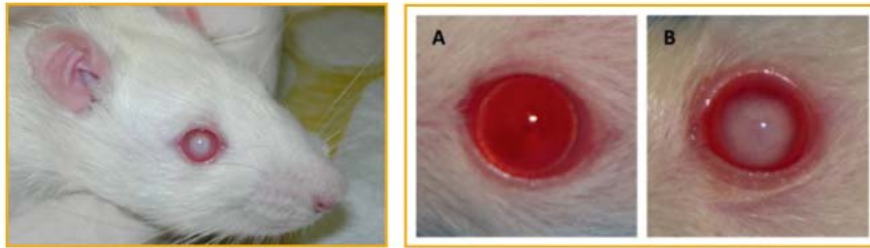
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Issues that relate to the lens

- Understand more about the protein localization in the ocular lens.
 - Interesting lens facts:
 - From “womb to tomb”
 - No protein turnover
 - Limited translated proteome
 - Expanded PTM proteome though!
 - Predominantly alpha crystallin proteins
 - small heat-shock proteins

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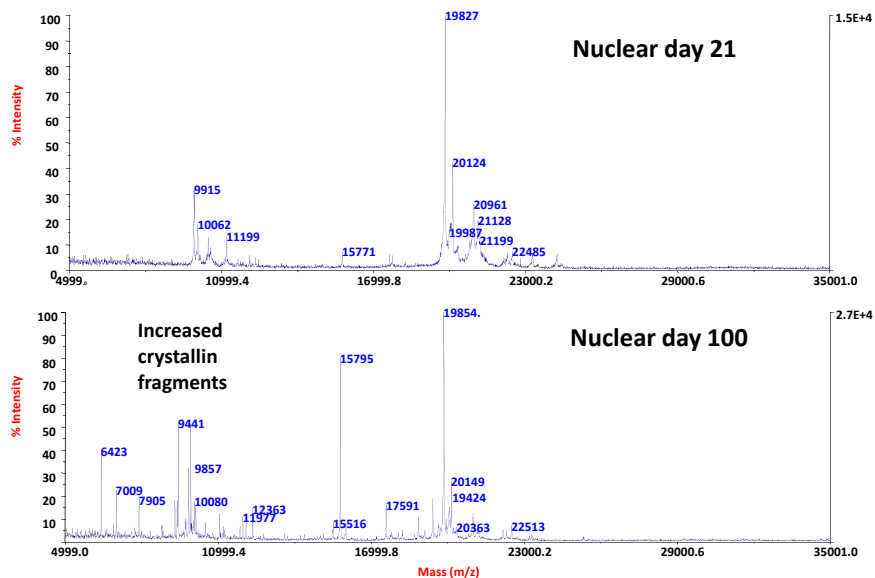
The rat model we use



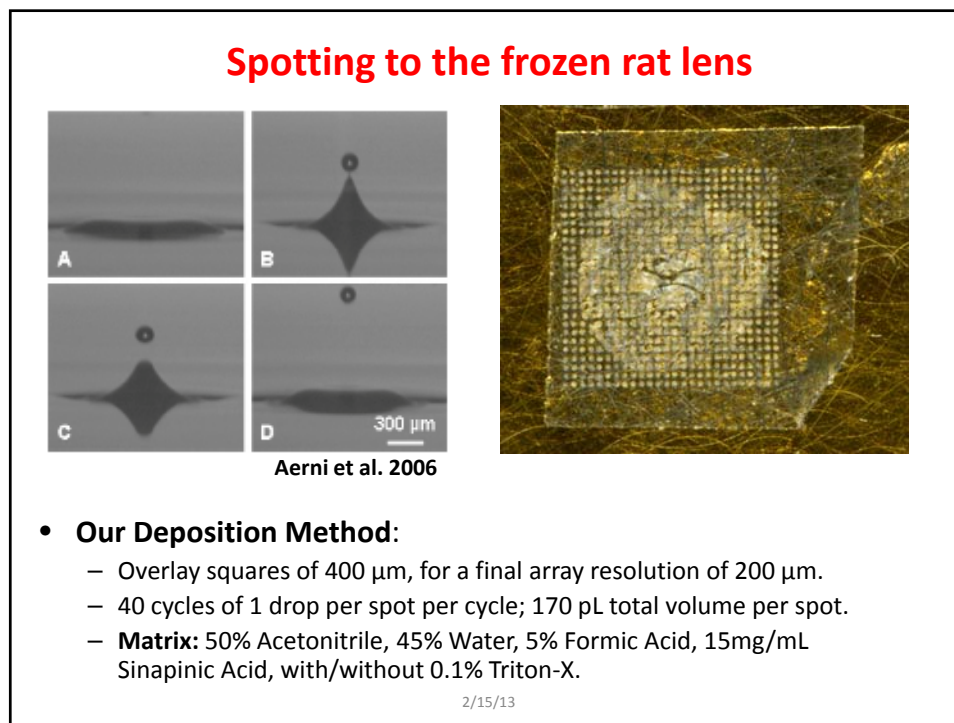
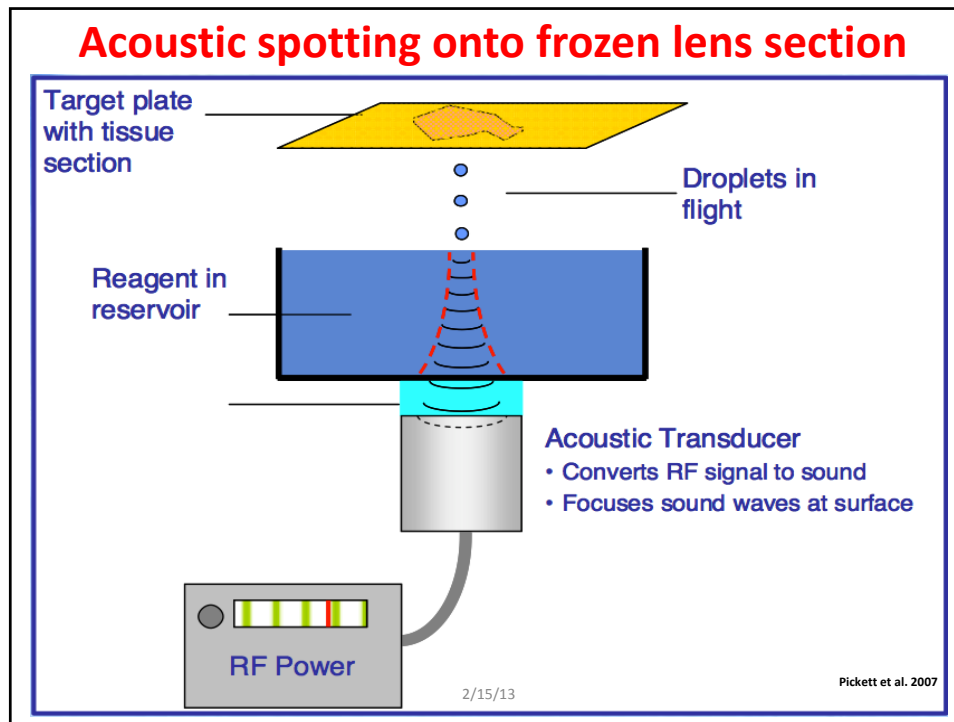
- ICR/f rat (Ihara/Inherited Cataract Rat, strain-f)
 - Model of age-related disease.
 - Spontaneously develops cataracts by 10 weeks of age.
 - Possible result of early oxidative insult.
 - Compare 21-day vs. 100-day

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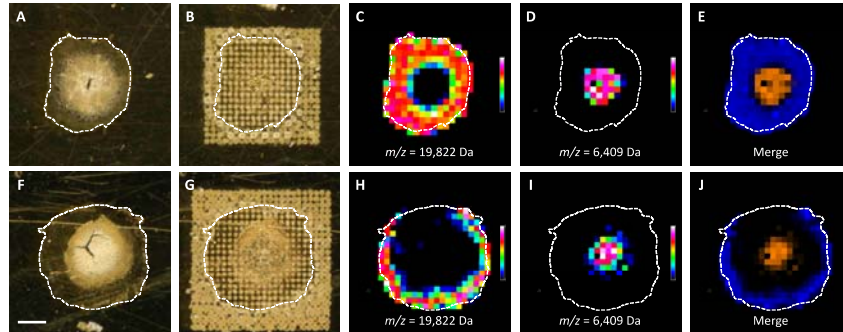
MALDI-TOF MS profiling of nuclear region of SD rat lens



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Lens Cataract: imaging reveals a geographic distribution of protein forms



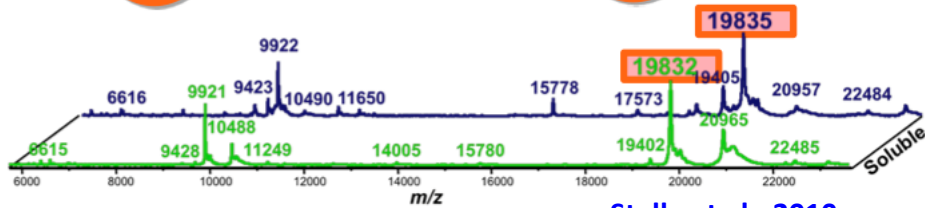
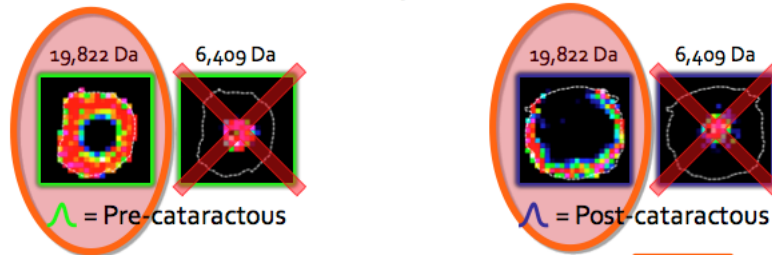
What are the proteins that imaging is detecting?

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Stella et al., 2010

Aqueous extract of the lens

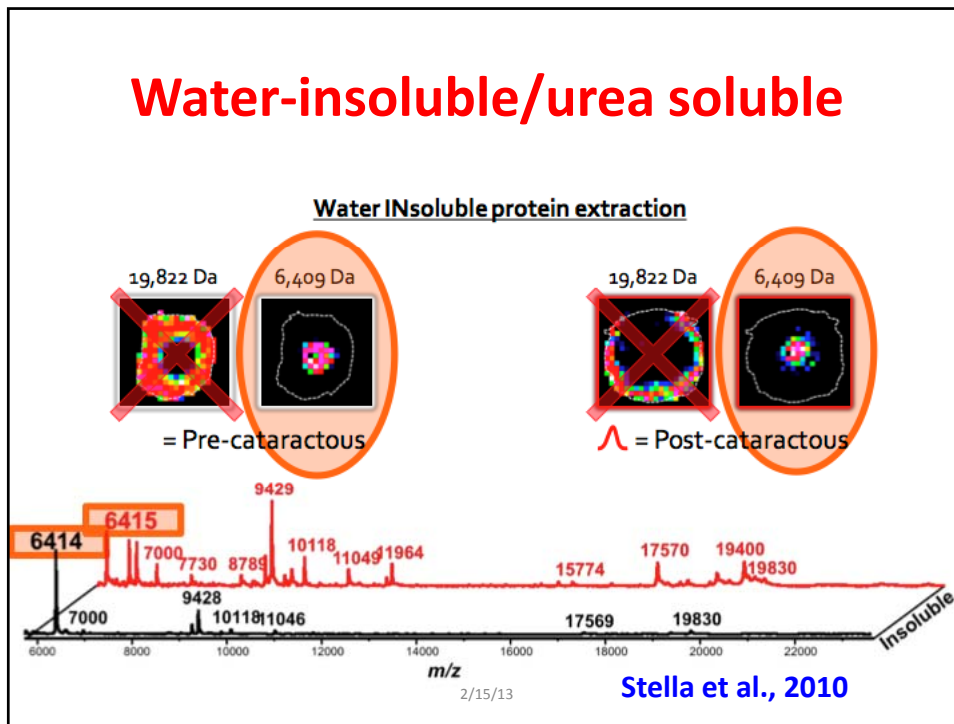
Water soluble protein extraction



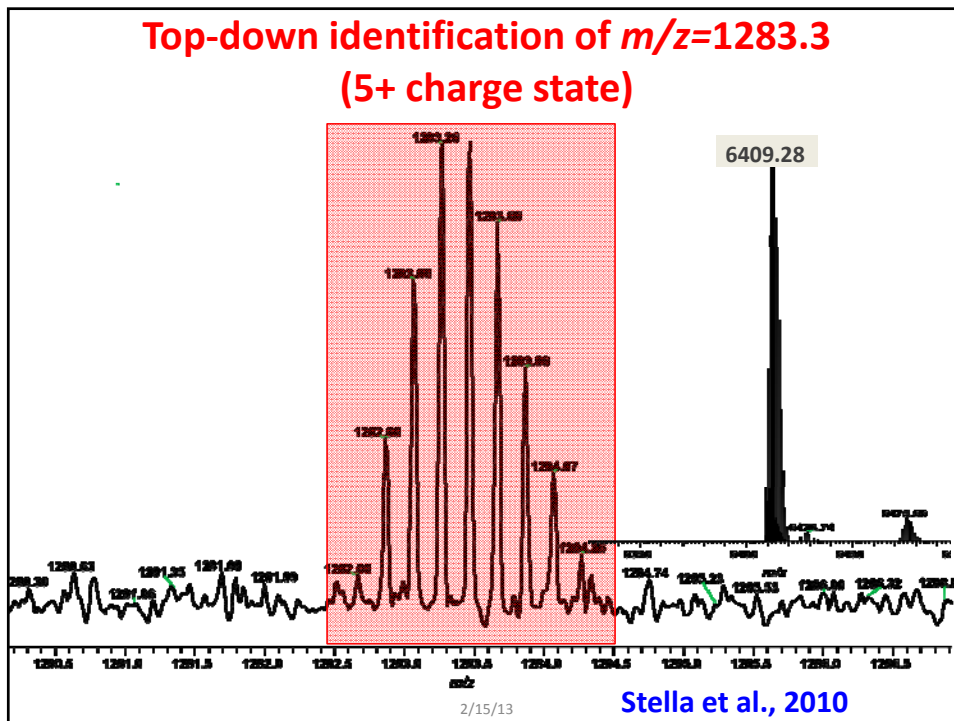
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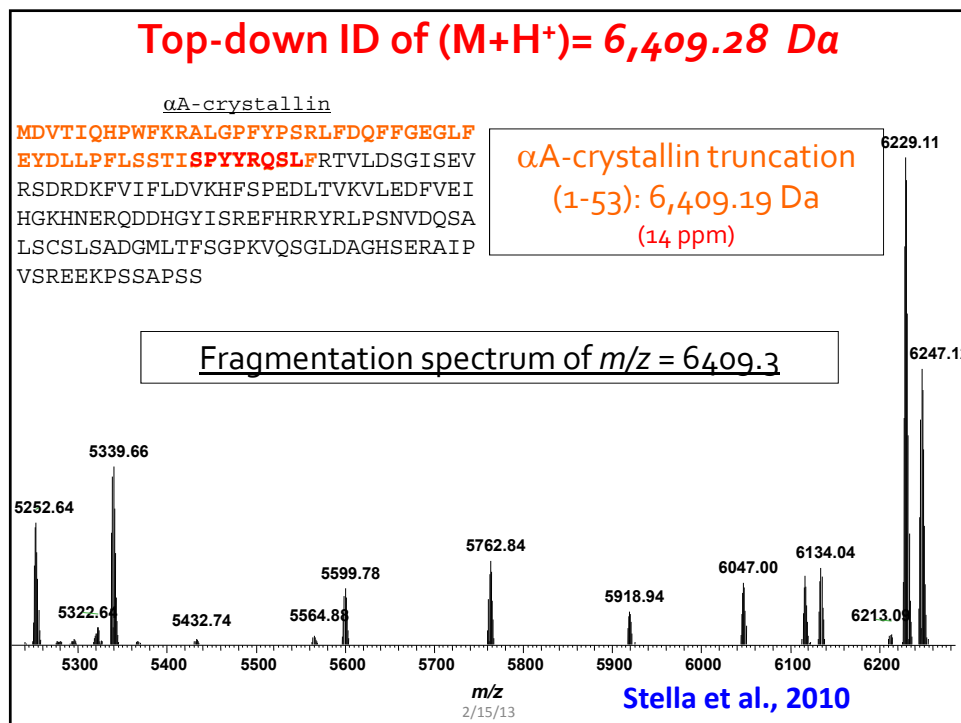
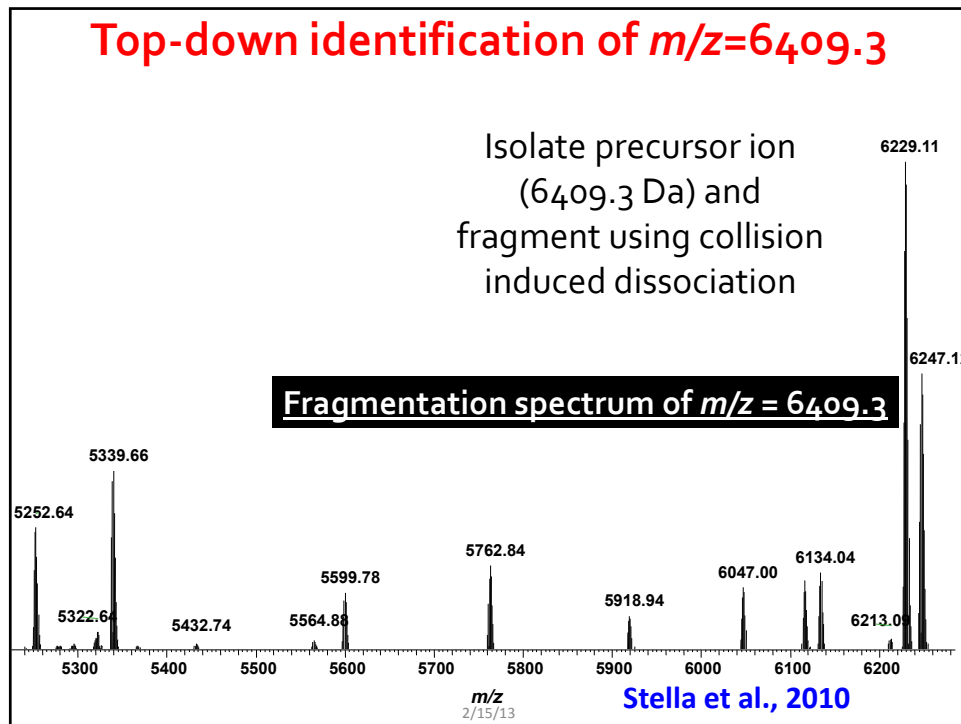
Stella et al., 2010

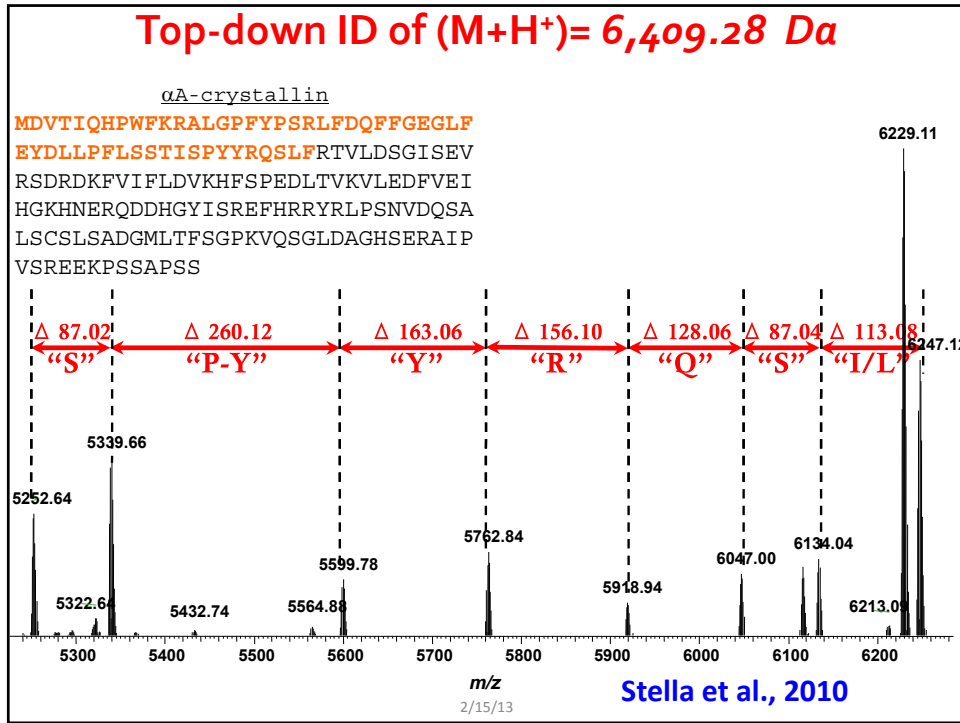
Water-insoluble/urea soluble



Top-down identification of $m/z=1283.3$ (5+ charge state)







Top-Down Protein Assignment

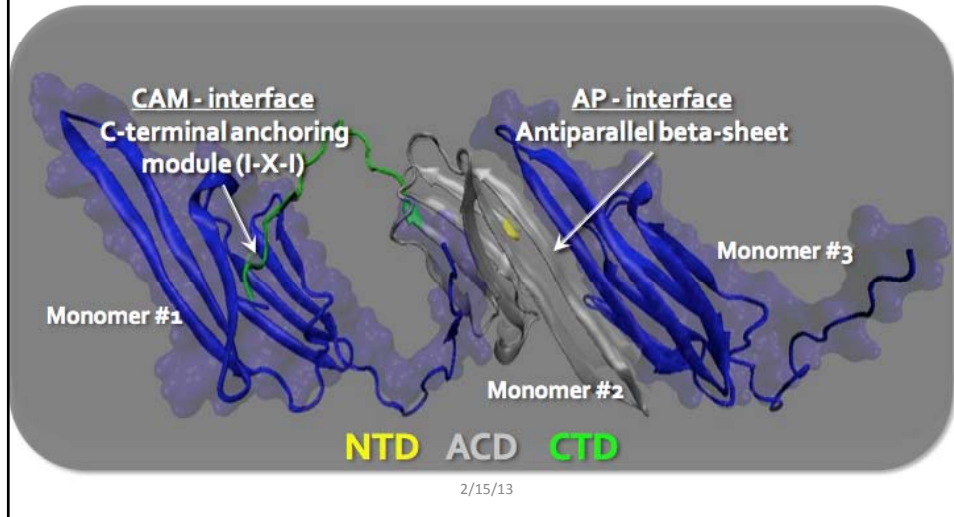
Protein ID	Predicted Mass (Da)*	Residues
Crystallin, alpha A	5,053.50	1-42
Crystallin, alpha A	6,409.19	1-53
Crystallin, alpha A	6,565.29	1-54
Crystallin, alpha A	9,284.76	1-78
Crystallin, alpha A	9,421.80	1-79
Crystallin, alpha A	10,110.11	1-85
Crystallin, alpha A	11,041.61	1-93
Crystallin, alpha A	11,842.04	1-100
Crystallin, alpha A	11,956.08	1-101
Crystallin, alpha A	17,562.77	1-151*
Crystallin, alpha A	18,043.96	1-156*
Crystallin, alpha A	18,200.06	1-157*
Crystallin, alpha A	18,823.44	1-163*
Crystallin, alpha A	19,393.70	1-168*
Crystallin, alpha A	19,822.89	1-173*

Full length →

* = N-terminal acetylation included (+42.01 Da)

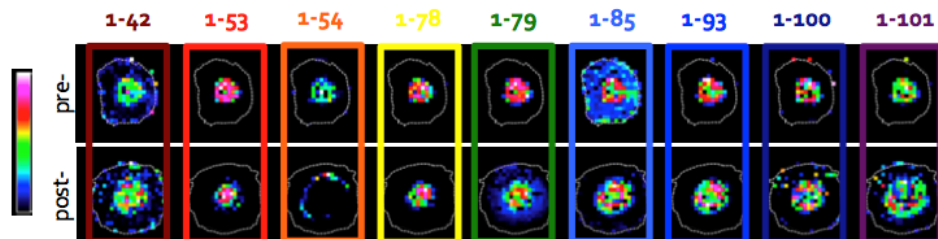
Stella et al. (2010) Invest Opth Vis Sci:5153 - 61.

Structure of α A-crystallin



Distribution of truncated α A-crystallins

1. Nuclear localization



MDVTIQHPWFKRALGPFYPSRLFDQFFGEGLEFYDLLPFLSSTI
 SPYYRQSLFRTVLDSGISEVRSRDKFVIFLDVKHFSPEDLTVK
 VLEDFVEIHGKHNERQDDHGYISREFHRRYRLPSNVDQSALSC
 SLSADGMLTFSGPKVQSGLDAGHSERAIPVSREEKPSSAPSS

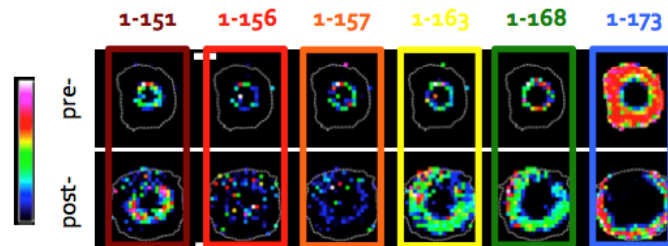
NTD ACD CTD

Stella et al., 2010

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Distribution of truncated α A-crystallins

2. Cortical/Nuclear-ring localization



MDVTIQHPWFKRALGPFYPSRLFDQFFGEGLF EYDLLPFLSSTI
 SPYYRQSLFRTVLDSGISEVRSRDKFVIFLDVKHFSPEDLTVK
 VLEDFVEIHGKHNERQDDHGYISREFHRRYRLPSNVDQSALSC
 SLSADGMLTFSGPKVQSGLDAGHSERAI PVSR EEEKPSSAPSS

NTD ACD CTD

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Stella et al., 2010

Histochemical imaging of concentric shells in lens

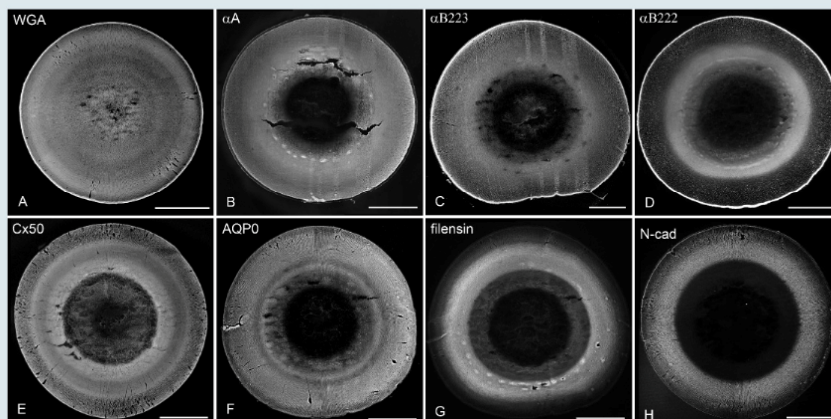


Figure 3(A-H): Composite of mouse lens sections labelled with wheat germ agglutinin (WGA) or antibodies to lens proteins. A) WGA (Invitrogen W32466); B) α A221 (Enzo ADI-SPA 221); C) α B223 (Enzo ADI-SPA 223); D) α B222 (Enzo ADI-SPA 222); E) connexin 50 (Cx50 from Dr. Thomas White); F) Aquaporin0 (AQP0; ADI-AQP02); G) filensin (Dr. Roy Quinlan 3241); H) N-cadherin (Ncad; BD 610920). Note the distinct patterns of shells for each label and the abrupt change from one shell to another. Magnification bars=500 μ

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Clark et al., ARVO, 2011

Optimization of spotting methods

Plate-Mounted, Vertical EtOH wash

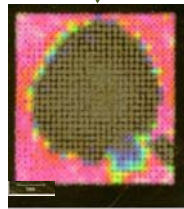
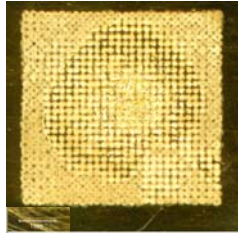
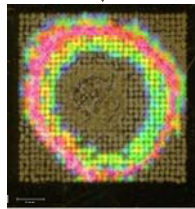
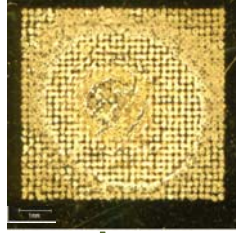
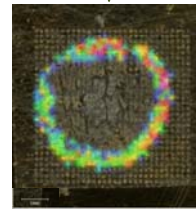
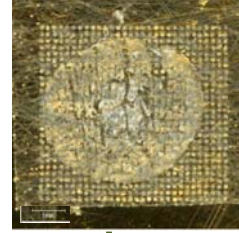


Plate-Mounted, Horizontal EtOH wash with Blotting

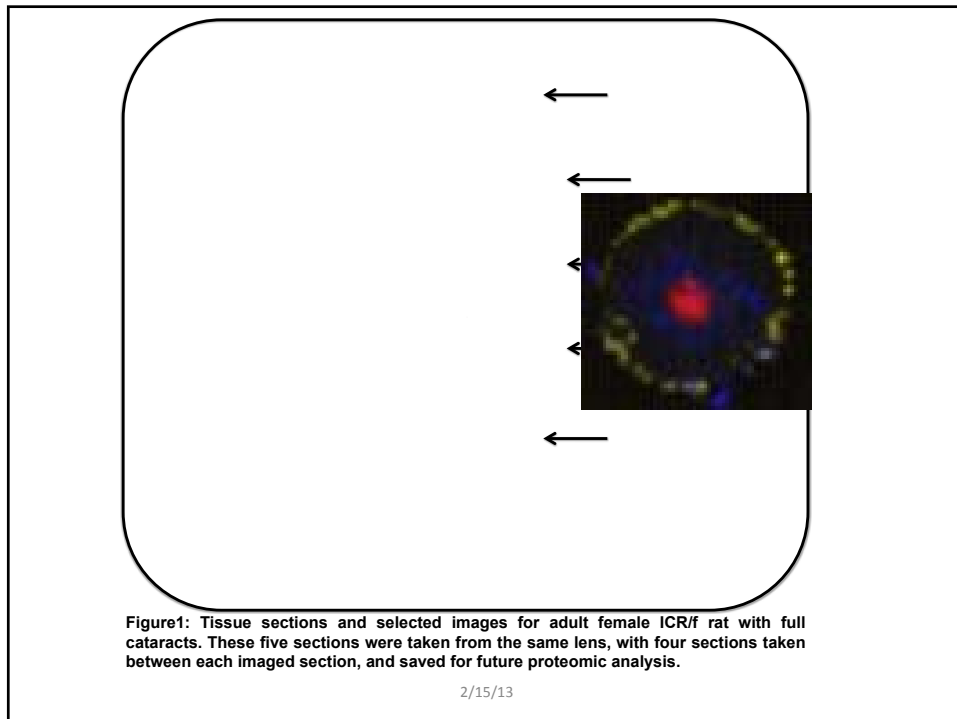


Carbon Fiber Tape Mounted and Lyophilized



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Slide adapted and modified from Kyle Floyd



Small molecule imaging

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Imaging of lipids in tissue

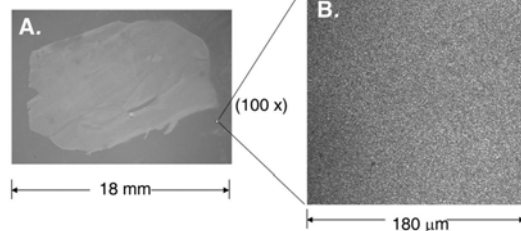


Fresh frozen tissue



Frozen tissue sectioning

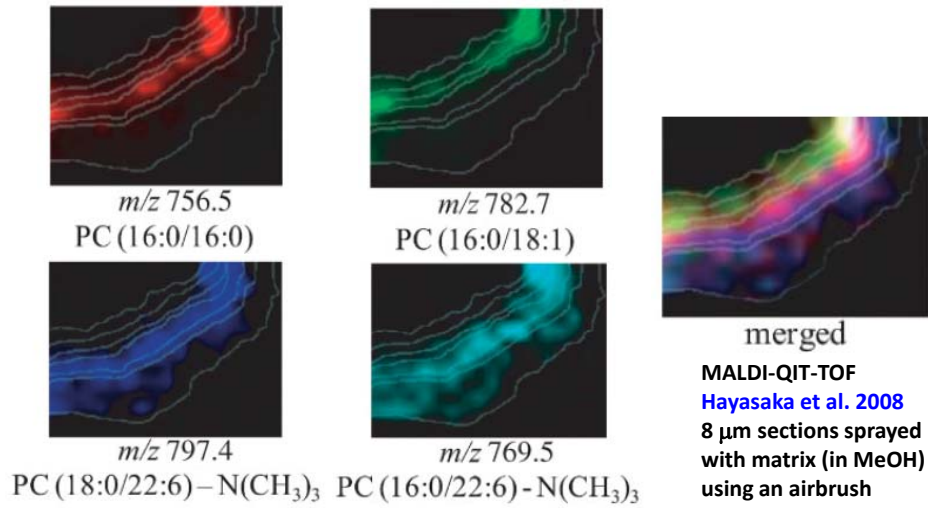
Cut and transfer 10-20 micron section to conductive glass slide



DHB (MALDI matrix) is transferred by sublimation.
This produces a very uniform coating (as shown in B) and high resolution images

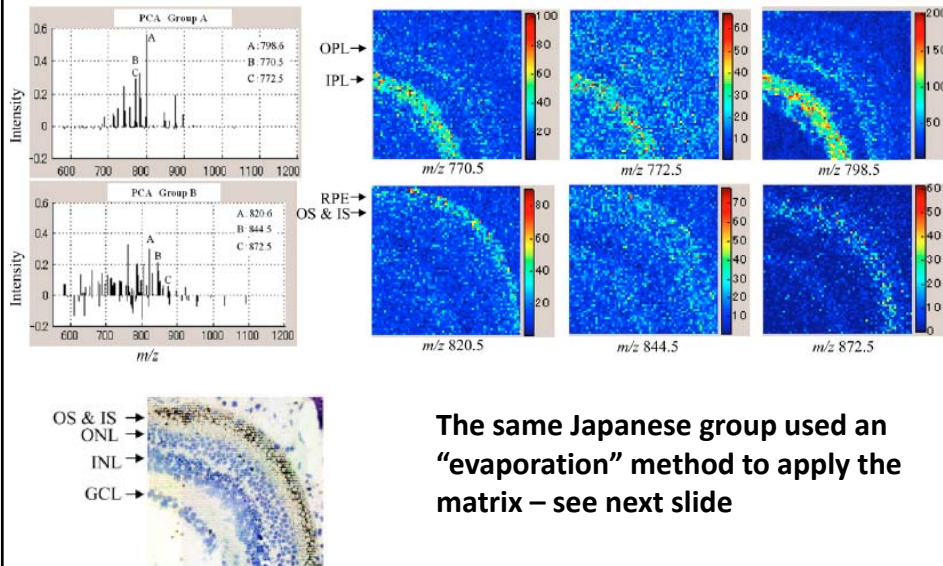
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Differential lipid distribution in the retina



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Using a higher resolution approach



The same Japanese group used an “evaporation” method to apply the matrix – see next slide

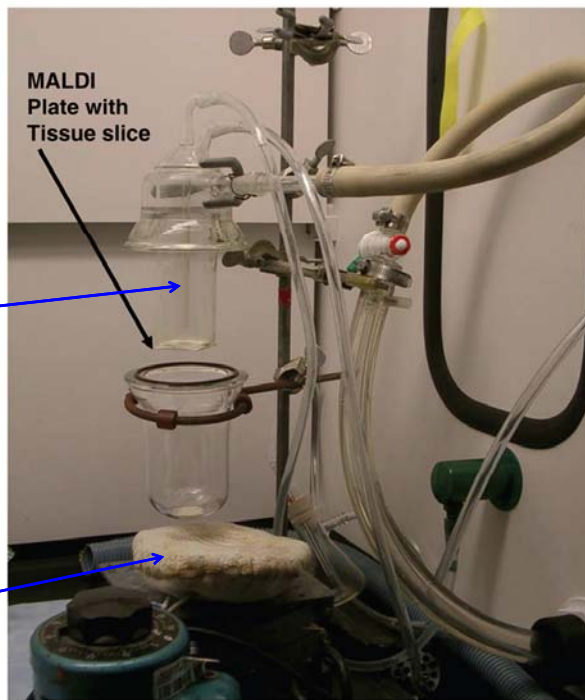
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Sublimation method for applying MALDI matrix to the tissue slice

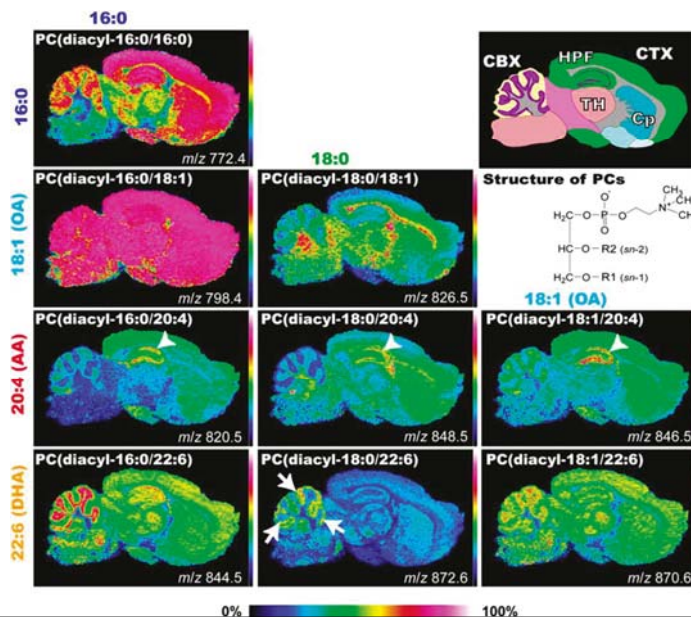
MALDI plate is upside down – ice is placed inside the glass “finger” to condense the DHB

A cold trap (not shown) is needed after the heated chamber to protect the vacuum pump

Sand bath to heat the DHB

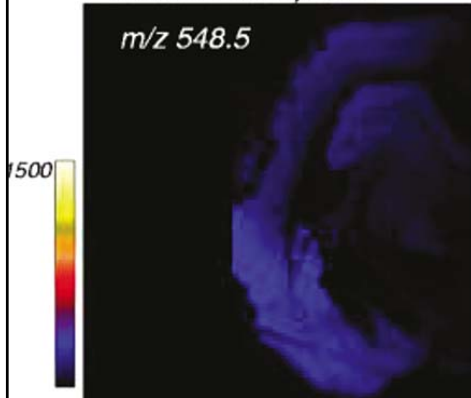


PC species distribution in the brain

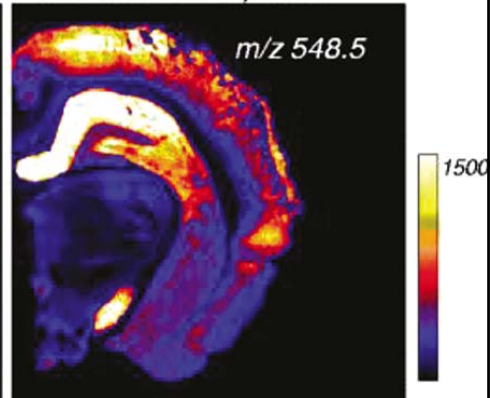


Ceramides and ischemia by IMS

(a) MALDI IMS
Control Hemisphere



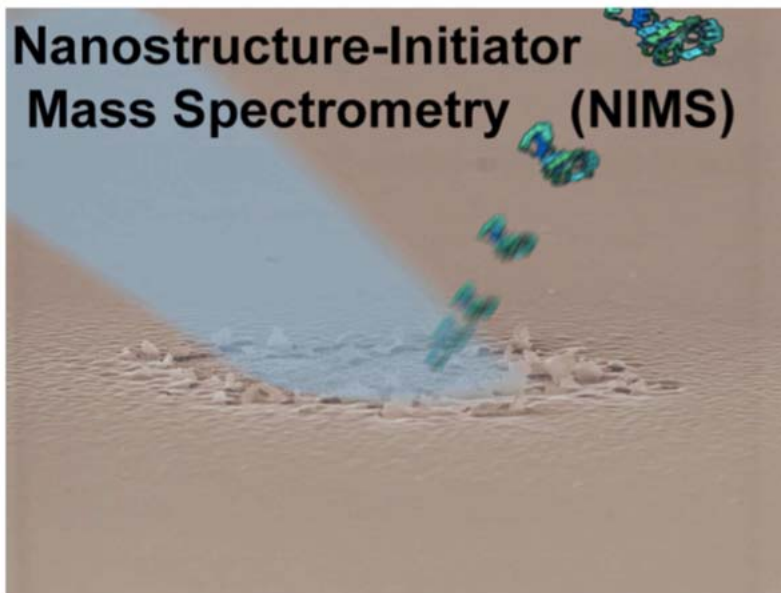
(b) MALDI IMS
Ischemic Hemisphere



The ion was determined to be Cer 18:0/18:1 (-H₂O)

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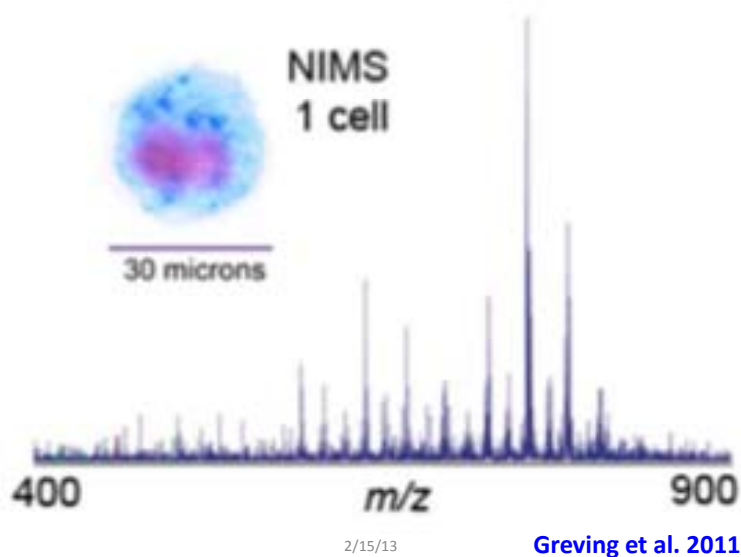
Nanostructure-Initiator Mass Spectrometry (NIMS)



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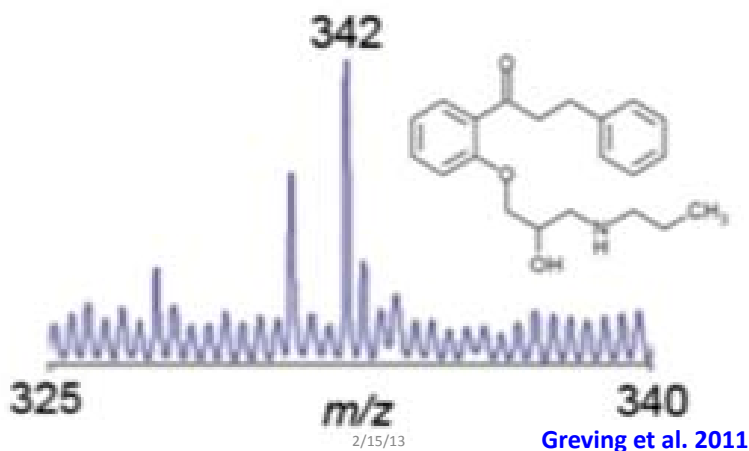
Greving et al. 2011

Spectrum from a single cell



Extreme sensitivity

propafenone (650 ymol)



Mass Spectrometry Imaging Workshop
April 20-24, 2013
Vanderbilt University

[https://www.msri.mc.vanderbilt.edu/
aims2013](https://www.msri.mc.vanderbilt.edu/aims2013)

Registration: \$750 before Mar 15
\$950 after Mar 15, up to Mar 31

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